

Role Of Fnac In Diagnosis Of Etiological Profile Of Lymphadenopathy

Dr. Bhawana Kumari¹, Dr. Manojit Midya¹, Dr. Deepti Sukheeja¹, Dr. Vinod Kumar Garg²

¹Assistant Professor, ²Associate Professor,

Govt. Medical College, Kota

***Corresponding Author:**

Dr. Vinod Kumar Garg

Associate Professor, Govt. Medical College, Kota,

Type of Publication: Original Research Paper

Conflicts of Interest: Nil

Abstract

Lymph nodes are common site of metastasis for different cancers. Thus the clinical recognition and urgent diagnosis of palpable lymphadenopathy is of paramount importance specially to differentiate between inflammatory lesions or metastatic or primary neoplastic tumor. Inclusion of FNAC in the diagnostic protocol as a first line non-invasive investigation seems to be quite useful to confirm or exclude- for example, metastatic disease – especially if a known history of malignancy is present. The results of FNAC compare favourably with those of tissue biopsies and in some situations the aspirate has qualities of a biopsy. Suspicious or doubtful situations should be resolved by surgical biopsy and further by immunohistochemistry and molecular techniques whenever required.

The aim of the present study is to highlight the role of FNAC in diagnosis of etiological profile of lymphadenopathy and to find out the accuracy of FNAC in comparison to biopsy.

Keywords:

Introduction

Lymphadenopathy is one of the commonest clinical presentation in clinical practice. There are approximately 800 lymph nodes in the body and no fewer than 300 of them lie in the neck. ⁽³⁸⁾

Lymphadenopathy is an abnormal increase in size and/ or altered consistency of lymph nodes. It is a clinical manifestation of regional or systemic diseases and serves as an excellent clue to the underlying disease. ⁽⁵⁰⁾

Enlargement of lymph node may result from the proliferation of lymphocytes intrinsic to lymph nodes, due to an infection or a lymphoproliferative disorder or from migration and infiltration of nodal tissue by either intrinsic inflammatory cells or metastatic malignant cells. ⁽⁵⁶⁾

Size of a node, to which some pathological significance can be attached, depends on the site. In

the cervical and axillary regions a lymph node > 1 cm, in the inguinal region

> 1.5 cm size, is considered significant. Causes of lymphadenopathy are reactive lymphadenitis, infections, malignant lymphoma and leukemias, metastatic malignancies and autoimmune disorder. It is important to differentiate between benign and malignant conditions to decide the further line of management.

The etiological profile varies from country to country and region to region. In developing countries like India, acute upper respiratory infections, suppurative skin lesions and tuberculosis are the major causes of regional lymphadenopathy. ⁽⁷⁷⁾ It has been stated that any significant lymph node enlargement not subsiding or remaining static in size for >2 weeks after conventional antibiotic therapy should be thoroughly investigated. ⁽¹⁴⁾

The diagnostic modalities that can be employed to the etiology of lymphadenopathy are FNAC and excision biopsy. FNAC is particularly helpful in the work-up of cervical masses and nodules because biopsy of cervical adenopathy should be avoided unless all other diagnostic modalities have failed to establish a diagnosis. ^(56, 48)

Aspiration of lymph nodes for diagnostic purpose was reported as early as 1904 by Grieg and Gray who used this procedure in the diagnosis of trypanosomiasis. ^(30, 56)

In 1921, Guthrie attempted to correlate lymph node aspiration cytology with various disease processes. ^(34, 56) FNAC is nowadays recognized as a rapid diagnostic technique because of its simplicity, early availability of results, cost effective procedure, minimal trauma, absence of complications and can be performed in any setting, with results usually available within 24 hours. ^(42, 39)

With the help of FNAC, preliminary diagnosis of a lesion with an adequate sample can be given within a few minutes when a quick staining and microscopic examination is performed on site. ⁽³³⁾

The diagnostic utility of FNAC of lymph nodes has been described. However, its role in the morphologic assessment of lymphadenopathies does not seem to have been completely established due to the belief by some pathologists that FNAC samples do not supply enough diagnostic material. ⁽²⁴⁾

In patients with an unknown clinical history of malignancy, a positive FNAC diagnosis may be the first indication of malignancy and be useful in furthering the clinical investigation to search for an occult neoplasm. Since a decision on conservative versus radical management depends on the type of tumor or non-tumorous process, it is of

paramount importance that the FNAC diagnosis of the lymphadenopathy be as accurate as possible. ⁽³³⁾

Diagnosis of lymphadenopathy depends mainly on excision of a gland and histopathological examination. For this, general anaesthesia and hospitalisation are required. Fine needle aspiration cytology, on the other hand, is free from these disadvantages and can safely be used as an alternative or complementary investigative technique. ⁽⁵⁶⁾

Excision biopsy which gives definitely the final diagnosis, but open biopsy before appropriate evaluation may be detrimental for a number of reasons. The patient may believe that his disease has been cured and he does not need further therapy, wound infection may delay definitive surgery, and biopsy incision if placed improperly, may compromise the adequacy of the neck dissection performed later. Excision biopsy takes more time than FNAC, delay in management of primary tumors may eliminate the best opportunity of its removal with metastatic nodes. Not all hospital can afford a full- fledged histopathology department.

Although open biopsy with histological examination of excised tissue still remains the gold standard for diagnosis of lymph node enlargement, yet FNAC has now become an integral part of the initial diagnosis and management of patients presenting with lymphadenopathy. This simple technique has gained wide acceptance since it offers a high degree of accuracy, leading itself to outpatient diagnosis and thus reducing the cost of hospitalization.

Aims And Objectives

1. To study the etiological profile of lymphadenopathy as per Fine Needle Aspiration Cytology.
2. To study the etiological profile of lymphadenopathy as per Histopathological Examination.
3. To find out the accuracy of Fine Needle Aspiration Cytology in the diagnosis of lymphadenopathy.

Material And Methods

Place of study: Department of pathology, SMS Medical College and attached hospitals, Jaipur, Rajasthan.

Study design: accepting α error 0.5 and power 80% for 79.1% sensitivity for lymphadenopathy (seed article), sample size was calculated 34, by study size version 2 statistical software.

Study duration: from September 2011 onwards

Sample size: Fine Needle Aspiration Cytology and histological study of minimum of 34 cases, upto maximum of 50 cases if possible.

Sampling technique: each and every case of lymphadenopathy coming for histopathological examination.

Inclusion Criteria

1. Patients with various age, irrespective of their sex having enlarged lymph nodes of surgical etiology at different parts of body.
2. Patients who need to be treated surgically and not medically after diagnosed by FNAC.

Exclusion Criteria

1. Patients treated medically after FNAC
2. Patients with inconclusive FNAC or histopathology results
3. Unfit for surgical procedures
4. Lymph nodes inaccessible to FNAC procedures.
5. This study will be carried on patients with any palpable enlarged lymph nodes.
6. The material for study will comprise of
 - a) Cytosmear made from material obtained by fine needle aspiration using a 23-25gauge needle and 10 ml syringe with the help of plunger.
 - b) Biopsy from surgically removed tissue.
 - c) Immunohistochemical marker study as and when required.

Staining Techniques

1. Staining of cytology smears:

The *May Grunwald Giemsa* staining techniques were used for the staining of cytology smears. The reagents which were used in the staining of cytology smears are: May Grunwald's stain, Giemsa stain, Methanol, Glycerol, Conical flask and Phosphate buffer(pH-6.8).

Preparation of the stains:

1. **Preparation of May Grunwald stain:** 0.3 g of powdered dye was weighed out and transferred to a conical flask of 200-250cc capacity. 100cc of methanol was added and the mixture was warmed to 50°C. The flask was then allowed to cool at room temperature and was shaken several times during the day. After standing for 24 hours the solution was filtered. It was then ready for use.

2. **Preparation of Giemsa solution:** 1 g of Giemsa powder was dissolved in 54ml of glycerol at 50⁰ C and after cooling mixed with 84ml of methanol GR and filtered.

Air dried smears were fixed in a jar of methanol for 5 minutes.

1. Fixed smears were stained as follows: Dry the film in the air.
2. Fix by immersing in a jar of methanol for 15-20 min.
3. Transfer the fixed films to staining jar containing May-Grunwald-stain freshly diluted with an equal volume of buffered water. (15 min)
4. Transfer them without washing to a jar containing Giemsa's stain freshly diluted with 9 volumes of buffered water, pH 6.8.(10-15 min)
5. Transfer the slides to a jar containing buffered water pH-6.8, rapidly wash in 3 or 4 changes of water, and finally allow to stand undisturbed in water for a short time (2-5 min.) for differentiation.
6. Stand the slides upright to dry.
7. Mounted by a rectangular cover glass using DPX mountant.

Inference of MGG staining:

1. Nuclei and basophilic cytoplasmic components appeared blue
2. Neutrophilic granules appeared lilac
3. Red cells were yellowish red.
4. Cytoplasm of neutrophil was pale pink

2. Haematoxylin & eosin staining technique

Paraffin sections were kept into water after the following procedure:

1. The paraffin of the section was melted over gentle flame.
2. Then the slides were kept in a jar of xylene for 5 minutes.
3. Again slides were kept in a Jar of xylene for 5 minutes.
4. Then sections were transferred to absolute alcohol for 5 minutes.
5. Then they were transferred to 80% alcohol.
6. Then were transferred to 50% alcohol.
7. Finally were washed in running water for 2 to 5 minutes.

8. Section were stained in: Meyer's haematoxylin solution for 5 minutes.
9. Rinsed in water and the slides were passed through gentle stream of running tap water.
10. Differentiated with 1 percent hydrochloric acid solution under microscopic checkup.
11. Counter stained with 1 percent solution of eosin for 30 sections.
12. Rinsed and dehydrated 3 times in acetone for 1 minute each time.
13. Mounted in DPX.

3. Ziehl–Neelsen staining technique:

1. Fix the dry smear in methanol for 5 min.
2. Wash with running tap water
3. Flood slide with hot Carbol Fuchsin for 5-7 min
4. Rinse with tap water
5. Decolorize with 20% H2SO4 till smear is decolorized

6. Rinse with tap water until smear takes light pink color
7. Flood slide with Methylene Blue for 2-3min
8. Rinse with tap water
9. Blot dry and mount with DPX

The results of all the three procedures were compared and evaluated taking histopathology as the gold standard.

Observation And Results

In present study, 34 cases of lymphadenopathy admitted in SMS Hospital, Jaipur, were studied. All cases were aspirated and smear prepared. Tissues were biopsied and cytological findings were compared with histopathological examination.

The diagnosis was consistent with histopathology in 32 cases and not consistent in 2 cases. Therefore, an accuracy rate of 88.89% was obtained.

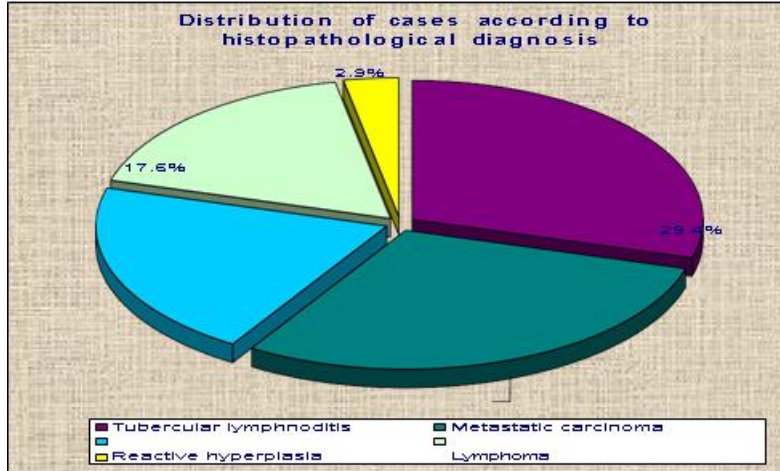
Clinicopathological Analysis:

In present study, commonest causes of lymphadenopathy were *Tubercular Lymphnoditis* (29.4%) and *Metastatic Carcinoma* (29.4%). Reactive Hyperplasia was (20.5%) at second place. Lymphoma constituted 17.6%, and Granulomatous Inflammatory lesion accounted for 2.94% of lymph nodes as shown in Table 1.

Table No. 01: Distribution of cases according to histopathological diagnosis

Histopathological Diagnosis	No of cases	Percentage (%)
Tubercular lymphnoditis	10	29.4%
Metastatic carcinoma	10	29.4%
Reactive hyperplasia	7	20.5%
Lymphoma	6	17.6%
Granulomatous Inflammatory lesion	1	2.94%
Total	34	100

Fig 1. Distribution of cases according to histopathological diagnosis

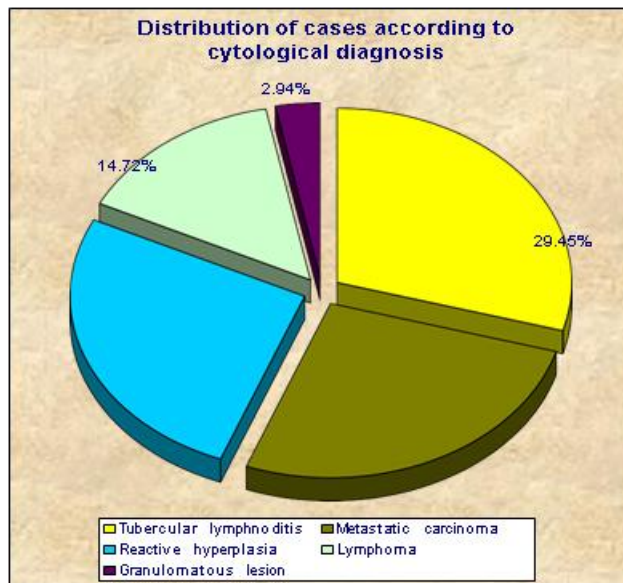


Out of 34 cases, cytological diagnosis was matched with histopathological diagnosis in 32 cases. One case which was reported as reactive hyperplasia by FNAC was found to be lymphoma on biopsy (Table 2). The other case which was reported reactive hyperplasia on FNAC, turned out to be metastatic carcinoma on histopathology.

Table No. 02: Distribution of cases according to cytological diagnosis

Cytological Diagnosis	No of Cases	Percentage
Tubercular lymphnoditis	10	29.4%
Metastatic carcinoma	9	26.4%
Reactive hyperplasia	9	26.4%
Lymphoma	5	14.7%
Granulomatous lesion	1	2.94%
Total	34	100%

Fig 2. Distribution of cases according to cytological diagnosis

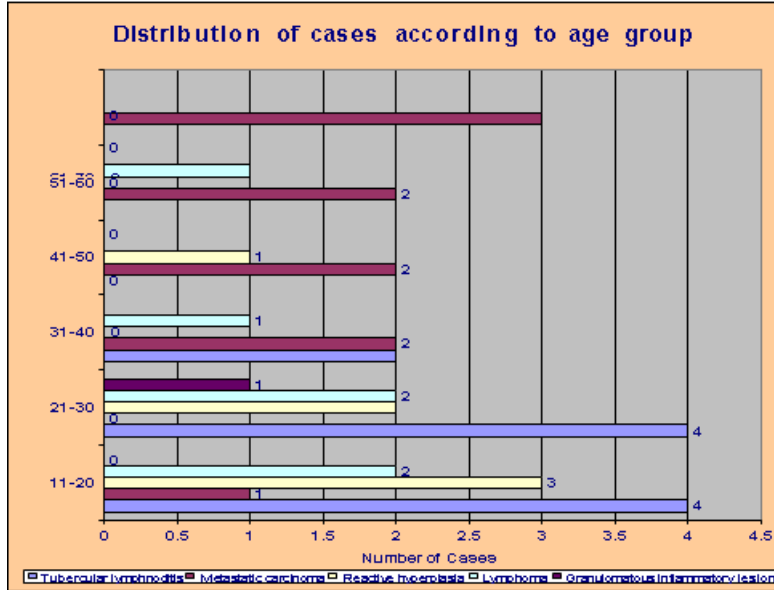


It showed that the 32.3% cases were in 1-20 yrs age group, 41.1% cases were between 21-40 yrs age group and 26.4% cases were in 41-70 years age group. Mean age was 32.67 years (Table 3).

Table No. 03: Distribution of cases according to age group

Age group (years)	Tubercular lymphnoditis	Metastatic carcinoma	Reactive hyperplasia	Lymphoma	Granulomatous inflammatory lesion	Total
1-10	-	-	1 (14.2%)	-	-	1
11-20	4 (40%)	1 (10%)	3 (42.8%)	2 (33.3%)	-	10
21-30	4 (40%)	-	2(28.5%)	2(33.3%)	1 (100%)	9
31-40	2 (20%)	2 (20%)	-	1(16.6%)	-	5
41-50	-	2 (20%)	1(14.2%)	-	-	3
51-60	-	2 (20%)	-	1(16.6%)	-	3
61-70	-	3 (30%)	-	-	-	3
Total	10	10	7	6	1	34

Fig 3. Distribution of cases according to age group



In metastatic carcinoma and lymphoma male preponderance was observed, while tubercular lymphnoditis and reactive hyperplasia show slight female preponderance. 55.8% cases were male and 44.1% cases were female. Male to Female ratio is 1.26:1.

Fig 4. Distribution of cases according to sex

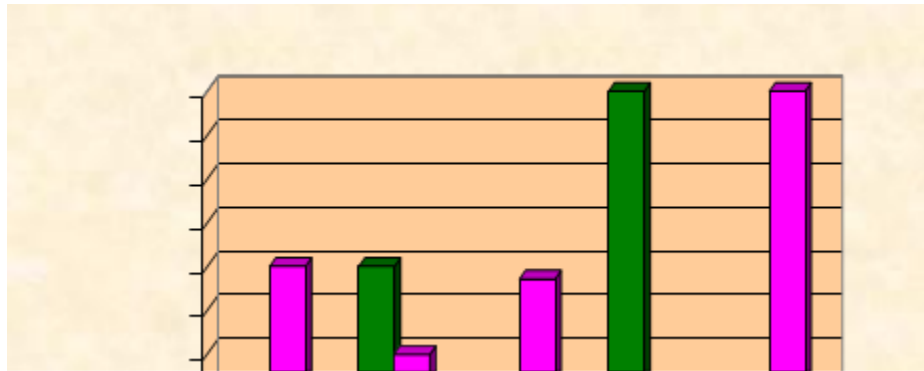


Table No. 04: Distribution of cases according to sex

Histological diagnosis	Male (%)	Female (%)
Tubercular lymphnoditis (n=10)	4 (40%)	6 (60%)
Metastatic carcinoma (n=10)	6 (60%)	4 (40%)
Reactive hyperplasia (n=7)	3 (42.8%)	4 (57.1%)
Lymphoma (n=6)	6 (100%)	-

Granulomatous inflammatory lesion (n=1)	-	1 (100%)

Correlation between cytological and clinical diagnosis was noted in 79.41% cases of lymphadenopathy. Out of 10 cases of Tubercular Lymphnoditis, 8 cases were diagnosed clinically as tuberculosis. In case of Metastatic Carcinoma, in 8 out of 9 cases clinically diagnosed were Metastatic Carcinoma. Out of 5 cases of Lymphoid neoplasm, clinical diagnosis was made in 4 cases. Accuracy of clinical diagnosis in relation to cytology was 79.41% as shown in Table 5.

Table No. 05: Co- relation between clinical diagnosis and cytological diagnosis

Clinical diagnosis	Cytological diagnosis					Total
	Tubercular lymphnoditis	Metastatic carcinoma	Reactive hyperplasia	Lymphoma	Granulomatous inflammatory lesion	
Tubercular lymphnoditis	8	1	2	-	-	11
Metastatic carcinoma	1	8	-	1	-	10
Reactive hyperplasia	1	-	6	-	-	7
Lymphoma	-	-	1	4	-	5
Granulomatous	-	-	-	-	1	1
inflammatory lesion						
total	10	9	9	5	1	34

Diagnosis in all 34 cases was confirmed by histopathology. Out of 34 cases, cytological diagnosis was matched with histopathological diagnosis in 32 cases. One case which was reported as reactive hyperplasia by FNAC was found to be Hodgkin lymphoma of mixed cellularity type on biopsy. The other case which was reported reactive hyperplasia on FNAC, turned out to be metastatic carcinoma on histopathology (Table 6).

Table No. 6: Co-relation between cytological diagnosis and final diagnosis

Histopathological diagnosis		Cytological diagnosis				
		Tubercular lymphnoditis	Metastatic carcinoma	Reactive hyperplasia	Lymphoma	Granulomatous inflammatory lesion
TB	10	10	-	-	-	-
Metastatic Ca	10	-	9	1	-	-
RH	7	-	-	7	-	-
Lymphoma	6	-	-	1	5	-
Granulomatous inflammatory lesion	1	-	-	-	-	1
Total	34	10	9	9	5	1

Sensitivity= 94.12%, Predictive value of positive test= 94.12%, Accuracy= 88.89%

P value was >0.05. However, FNAC was well correlated with histopathology, but due to less number of cases studied, it could not be proved to be statistically significant.

Table No. 07: Correlation of cytologic and histopathological diagnosis in cases of Metastatic Carcinoma

		Histopathological diagnosis		
		Positive	Negative	Total
Cytological diagnosis	Positive	9 (true positive)	0 (false positive)	9
	Negative	1(false negative)	0 (true negative)	1
	Total	10	0	10

Sensitivity =90%, Accuracy =90%, Specificity=100% and Predictive value of positive test=100%

Table No. 08: Corelation of cytologic and histopathological diagnosis in cases of lymphoma

		Histopathological diagnosis		
		Positive	Negative	Total
Cytological diagnosis	Positive	5 (true positive)	0 (false positive)	5

	Negative	1 (false negative)	0 (true negative)	1
	Total	6	0	6

Sensitivity =83.33%, Specificity=100%, Predictive value of positive test=100%, Accuracy =83.33

Out 9 cases of reactive hyperplasia, 7 cases (77.7%) were correlated with finding of histopathology. 2 cases were false positive by FNAC, one was diagnosed as lymphoma and other one as metastatic carcinoma in final diagnosis.

Table No. 09: Corelation of cytologic and histopathological diagnosis in cases of reactive hyperplasia.

		Histopathological diagnosis		
		Positive	Negative	Total
Cytological diagnosis	Positive	7 (True Positive)	2 (False Positive)	9
	Negative	0 (False Negative)	0 (True Negative)	0
Total		7	2	9

Sensitivity =100%, Predictive value of positive test = 77.78%, Accuracy = 77.78%

Table No. 10: Corelation of cytologic and histopathological diagnosis in cases of tuberculosis and granulomatous inflammatory lesion

Statistical values	Tubercular lymphnoditis	Granulomatous inflammatory lesion
True positive	10	1
True negative	0	0
False positive	0	0
False negative	0	0
Total	10	1

Sensitivity, Specificity, predictive value of positive test of FNAC in tubercular lymphnoditis and granulomatous inflammatory lesion was 100% and diagnostic accuracy also as 100%

Summary And Conclusion

Lymphadenopathy is one of the common clinical presentation of various ongoing disease process

inside the body. The present study was conducted to find out etiological profile of lymphadenopathy as per cytological and histological examination and to evaluate the accuracy of FNAC in relation to histopathology.

The recent trend in medical practice is toward adopting a diagnostic modality that is both cost effective and minimally invasive. In this regard the diagnostic utility of FNAC seems to be ideal as a first line investigation in evaluating patients presenting with lymphadenopathy. FNAC is a reliable diagnostic method of providing with ease the identification of metastasis in patients with a suspicion of or known malignancy apart from other process.

Fine needle aspiration of all these lesions was done. All cytosmears were stained with hematoxylin and eosin and May Grunwald Giemsa stains. The cytomorphological features were recorded.

The aim of the present study is to highlight the role of FNAC in diagnosis of etiological profile of lymphadenopathy and to find out the accuracy of FNAC in comparison to biopsy. In our study a total of 34 patients were taken for study.

Subsequently the lymph nodes were excised or an open biopsy done and paraffin sections were prepared which were stained with routine hematoxylin and eosin stains. The results of cytomorphology were then correlated with histopathology. Out of 34 cases, the diagnosis was consistent with histopathology in 32 cases and not consistent in 2 cases. Therefore an accuracy rate of 88.89% was obtained and summarised as:

1. Cases of lymphadenopathy showed a wide range of age from 10 to 65 years. Mean age was 32.67 years.
2. Tubercular Lymphnoditis and Metastatic Carcinoma were the commonest causes in our study.
3. Commonest cause of lymphadenopathy in children was Reactive Hyperplasia, in adults Tubercular Lymphnoditis and Lymphoma and in older age Metastatic Carcinoma.
4. Immunohistochemistry was performed to know the type of lymphoma and to confirm the diagnosis. Two cases (33.3%) on IHC were found as Hodgkin lymphoma and 4 cases (66.7%) as non Hodgkin Lymphoma.

5. In our study, overall Sensitivity of FNAC was 94.12% and Diagnostic accuracy as 88.89%. Thus, FNAC was well correlated with histopathology.
6. FNAC is a simple and very accurate technique for the diagnosis of lymphadenopathy and may be used as a first line investigation in lymphadenopathy as a screening tool in outpatient clinic.

References

1. Advani S K, Aqil S, Das J R, Dahar A. Role of Fine Needle Aspiration Cytology in the cervical lymphadenopathy. *Pak J Otolaryngol* 2008; 24(3): 42-4.
2. Agarwal D., Bansal P., Rani B., Sharma S., Chawla S., Bharat V., Sharma S.: Evaluation of etiology of lymphadenopathy in different age groups using Fine Needle Aspiration Cytology: A retrospective study. *The Internet Journal of Pathology*. 2010 Volume 10 Number 2.
3. Anderson JR, Armitage JO, Weisenberger DD (1998). Epidemiology of non Hodgkin's lymphomas: distributions of the major subtypes differ by geographical locations. *Non-Hodgkin's Lymphoma Classification Project*. *Ann Oncol* 9:717-720.
4. Annam V, Kulkarni MH, Puranik RB. Clinicopathologic profile of significant cervical lymphadenopathy in children aged 1-12 years. *Acta Cytologica*. 2009 Mar-Apr;53(2):174-8.
5. Anon (1997). A clinical evaluation the International Lymphoma Study Group Classification of non-Hodgkin's Lymphoma. *The Non-Hodgkin's Lymphoma Classification Project*. *Blood* 89:3909-3918.
6. Anon (2008). The International T-cell Lymphoma Project: International Peripheral T-cell and NK/T-Cell Lymphoma Study: Pathology findings and clinical outcomes. *J Clin Oncol*. In Press.
7. Armitage JO, Weisenberger DD (1998). New approach to classifying non- Hodgkin's lymphoma: clinical features of the major histologic subtypes. *Non Hodgkin's Lymphoma Classification Project*. *J Clin Oncol* 16: 2780-2795.
8. Bloch M., Comparative study of lymph node cytology in puncture and histopathology. *Acta Cytologica*, 1967, Vol. II, No. 2, 139.

9. Chan J K, Tsang W Y 1996 Reactive lymphadenopathy. In: Weiss L M (ed) Pathology of lymph nodes. Contemporary issue in surgical pathology. Vol. 21. Churchill livingstone, New York, 81-167
10. Chhotray GP, Acharya GS. Fine needle aspiration cytology in diagnosis of metastatic lymphadenopathies .Indian J Med Res 1987; 85: 685-688.
11. Collins BT, Elmberger PG, Tani EM, et al. Fine needle aspiration of Merkel cell carcinoma of the skin with cytomorphology and immunocytochemical correlation. Diagn Cytopathol 1998;18:251-57.
12. Dandapat M.C, Panda B.K., Patra A.K. and Acharya N.; Diagnosis of tubercular lymphadenitis by Fine Needle Aspiration Cytology; Ind. J. Tub., 1987, 34, 139
13. Dandapat M. C. *, Mishra B. M., Dash S. P., Kar P. K; Peripheral lymph node tuberculosis: A review of 80 cases; British Journal of Surgery; Volume 77, Issue 8, pages 911–912, August 1990
14. De Las Casas LE, Gokden M, Mukunyadzi P, et al. A morphological and statistical comparative study of small cell carcinoma and non Hodgkin lymphoma in fine needle aspiration biopsy material from lymph nodes. Diagn Cytopathol 2004;31:229-34.
15. Dey P, Jogai S, Amir T, et al. Fine needle aspiration cytology of Merkel cell carcinoma. Diagn Cytopathol 2004;31:364-65.
16. Drukar BJ, Talpaz M, Resta DJ, Peng B, Buchdunger E, Ford JM, Lydon NB, Kantarjian H, Capdeville R, Ohno-Jones S, Sawyers CL (2001). Efficacy and safety of a specific inhibitor of the BCR-ABL tyrosine kinase in chronic myeloid leukemia. N Engl J Med 344:1031-1037.
17. Farooq A, Ameen I. Comparison FNAC vs excision biopsy for suspected tuberculous cervical lymphadenopathy. Ann King Edward Med Coll 2003; 9(3): 21608
18. Ferrer R, et al: Lymphadenopathy: Differential diagnosis and evaluation. Am Fam Physician 58:1313, 1998 [PMID: 9803196]
19. Frable WJ. Thin needle aspiration biopsy: A personal experience with 469 cases. Am J Clin Pathol 1976; 65: 168-181
20. Gupta S.K. Dutta T.K. and Aikat B.K. Lymph node aspiration biopsy in diagnosis of lymphoma. Ind. J. Path, and Microbiol. 1977,20, 231.
21. Haque MA, Talukder SI. Evaluation of fine needle aspiration cytology (FNAC) of lymph node in Mymensingh. Mymensingh Med J 2003; 12: 33-35
22. Harris N L, Jeffe E S, Stein H et al. 1994. A revised European-American classification of lymphoid neoplasm: a proposal from the International Lymphoma Study Group. Blood 84: 1361-1392.
23. Hirachand S, Lakhey M, Akhter J, Thapa B, Evaluation of fine needle aspiration cytology of lymph nodes in Kathmandu Medical College, Teaching hospital, Kathmandu University Medical Journal (2009), Vol. 7, No. 2, Issue 26, 139-142
24. Howard D J, Lund V J. Pharynx, Larynx and neck. In: Williams N S, Bulstrode C J K, O'Connell P R, editors. Bailey and Love's Short Practice of Surgery. 25th ed. London: Edward Arnold (Publishers) Ltd; 2008; 702- 33.
25. Jan Muhamad Menon, Altaf Ahmed Talpur, Roshan A S, Abdul R Q. An audit of peripheral lymphadenopathy. Pakistan Journal of surgery 2007 Vol. 23 (3):183-186
26. Jiménez-Heffernan JA, Vicandi B, López-Ferrer P, Hardisson D, Viguier JM; Value of fine needle aspiration cytology in the initial diagnosis of Hodgkin's disease. Analysis of 188 cases with an emphasis on diagnostic pitfalls. Acta Cytol. 2001 May-Jun;45(3):300-6.
27. Khajuria R, Goswami K C, Singh K, Dubey VK. Pattern of Lymphadenopathy on Fine Needle Aspiration Cytology in Jammu. JK Science 2006; Vol. 8 (3): 157-159.
28. Khurana KK, Stanley MW, Powers CN, Pitman MB. Aspiration cytology of malignant neoplasms associated with granulomas and granulomas-like features. Cancer Cytopathol 1998; 84: 84-91
29. Kline TS, Kannan V, Kline IK. Lymphadenopathy and aspiration biopsy cytology, review of 376 superficial nodes. Cancer 54: 1076-1081, 1984
30. Kolte SS, Satarkar RN, Mane PM. Microfilaria concomitant with metastatic deposits of

- adenocarcinoma in lymph node fine needle aspiration cytology: A chance finding. *J Cytol* 2010; 27:7-80.
31. Kumar Arun , Nayar Mohini Chandra Mithilesh, critical appraisal of fine needle aspiration cytology in tuberculous lymphadenitis; *Acta Cytologica* 1992;36(3): 391-394.
 32. Layfield LJ. Fine-needle aspiration of the head and neck. *Pathology (Phila)* 1996;4:409–38.
 33. Malik G A, Rehan T M, Bhatti S Z. Relative frequency of diseases in patients with lymphadenopathy. *Park J Surg* 2003; 19(2):86-9.
 34. Morton LM, Wang SS, Devesa SS, Hatge P, Weisenberger DD, Linet MS (2006). Lymphoma incidence pattern by WHO subtype in the United States, 1992-2001. *Blood* 107: 265-276.
 35. Nada al Alwan, Al Hashimi AS, Salman MM, Al Attar EA. Fine needle aspiration cytology versus histopathology in diagnosing lymph node lesion. *Eastern Mediterranean Health Journal* 1996; 2: 320-325
 36. Nagpal B.L., Dhar C.N., Singh A. & Bahi H.H. Evaluation of imprint cytodiagnosis in case of lymphadenopathy. *Ind. J.P.M.*, 1982, 25, 35.
 37. Narang R.K., S. Pradhan, Singh R.P. And Chaturvedi S. Place of fine needle aspiration cytology in the diagnosis of lymphadenopathy. *Ind. J. Tub.*, 1990, 37, 29
 38. Ojo BA, Buhari MO, Malami SA, Abdul Rahaman MB. Surgical lymph node biopsies in University of Ilorin Teaching Hospital, Ilorin, Nigeria. *Niger Postgrad Med J* 2005; 12: 299-230
 39. Orell SR, Sterett GF, Whitaker D, van Heerde P, Miliuskas J. Lymph nodes. Fine needle aspiration cytology. 4th ed. Elsevier: New Delhi; 2005. Pp 81-124
 40. Pahwa R, Hedau S, Jain S, et al. assessment of possible tuberculous lymphadenopathy by PCR compared to non- molecular methods. *J Med Microbiol* 2005;54:873-78.
 41. Pilloti S, Di Palma S, Alasio L Bartoli C, Rilke F. Diagnostic assessment of enlarged superficial lymph nodes by aspiration biopsy. *Acta Cytol* 1993; 93: 853 -866.
 42. Prasad R R A, Narasimhan, Sankaran V, Veliath A J. Fine needle aspiration cytology in the diagnosis of superficial lymphadenopathy: An analysis of 2,418 cases. *Diagn Cytopathol* 1996; 15: 382-6.
 43. Rock A. Committa B: Lymphadenopathy. In *Textbook of paediatrics*. Seventeenth edition. Edited by W Nelson. RE Behraman. RM Kliegman. Philadelphia. WB Saunders, 1996, p 1441
 44. Sandanah MM, Gita J: Acid fast bacilli in aspiration smears from tuberculous lymph nodes; *Acta Cytologica* 1987;31 (11):17-19.
 45. Schoot Vande, Aronson D-c, Behrendt-h, Bras-J. The role of FNAC in children with superficial lymphadenopathy. *J Pediatric Surg* 2001; 15 (12): 7-11
 46. Schwarzbertt Marija Bizak: cyto morphological characteristic of non- Hodgkin's lymphoma; *Acta Cytologica* 1988; 32 (2): 216-220.
 47. Sheahan P, Fitzgibbon J, O'Leary G, Lee G. Efficacy and pitfalls of fine needle aspiration in the diagnosis of neck masses. *Surg J.R. Coll Surg Edinb Irel* 2004: 152-156
 48. Skoog L, Schmitt FC, Tani E. Neuroendocrine (Merkel cell) carcinoma of the skin: immunocytochemical and cytomorphological analysis on fine needle aspirates. *Diagn Cytopathol* 1990;6:53-57.
 49. Steel BJ, Schwartz MR, Ramzy I. Fine needle aspiration biopsy in the diagnosis of lymphadenopathy in 1103 patients. *Acta cytol* 1993; 39: 76- 81
 50. Stewart BW, Kleihues P (2003). *World Cancer Report*. IARC Press.
 51. Swerdlow S.H., Campo E., Haris N.L., Jaffe E.S., Pileri S.A., Stein H., Thiele J., Vardiman J.W., WHO classification of tumours of haematopoietic and lymphoid tissues; IARC: Lyon 2008
 52. Tariq Ahmad, Mohammad Naeem, Siddique Ahmad, Ambreen Samad. Fine Needle Aspiration Cytology (Fnac) And Neck Swellings In The Surgical Outpatient, *J Ayub Med Coll Abbottabad* 2008; 20(3): 30-32
 53. Tripathi S.N., Mishra N., Patel N.M., Samantray D.K., Das B.K., and Mania R.N., Place of aspiration biopsy in the diagnosis of lymphadenopathy, *Ind. J. Tuberc.* 1985,32,130.

54. Twist C J, Link M P: assessment of lymphadenopathy in children. *Pediatr Clin North Am* 2002;49:1009-1025
55. Udani P M: Tuberculosis in children. *Ind J Pediatr* 1994;61:451-462
56. Ustun M, Risberg B, Davidson B, et al. Cystic change in metastatic lymph nodes: a common diagnostic pitfall in fine needle aspiration cytology. *Diagn Cytopathol* 2002;27:387-92.
57. Wani BN, Jajoo SN. Ipsilateral axillary tubercular lymphadenopathy, contralateral osseous tuberculosis in a case of ductal carcinoma of breast. *Indian J Cancer* 2008; 45:182-4.
58. Warnke R A. Weiss L M. Chan J K et al. 1995 Tumors of the lymph nodes and spleen. Atlas of tumor pathology, 3rd series, Fascicle 14. Armed force institute of pathology, Washington DC
59. Winifred G, Gabrijela K, Diagnostic Cytopathology. Elsevier Health Science; 21 June 2010,3rd ed, 411-437.
60. Zajicek J. Aspiration biopsy cytology. Part I. Cytology of supradiaphragmatic organs. Basel: Karger;1974.